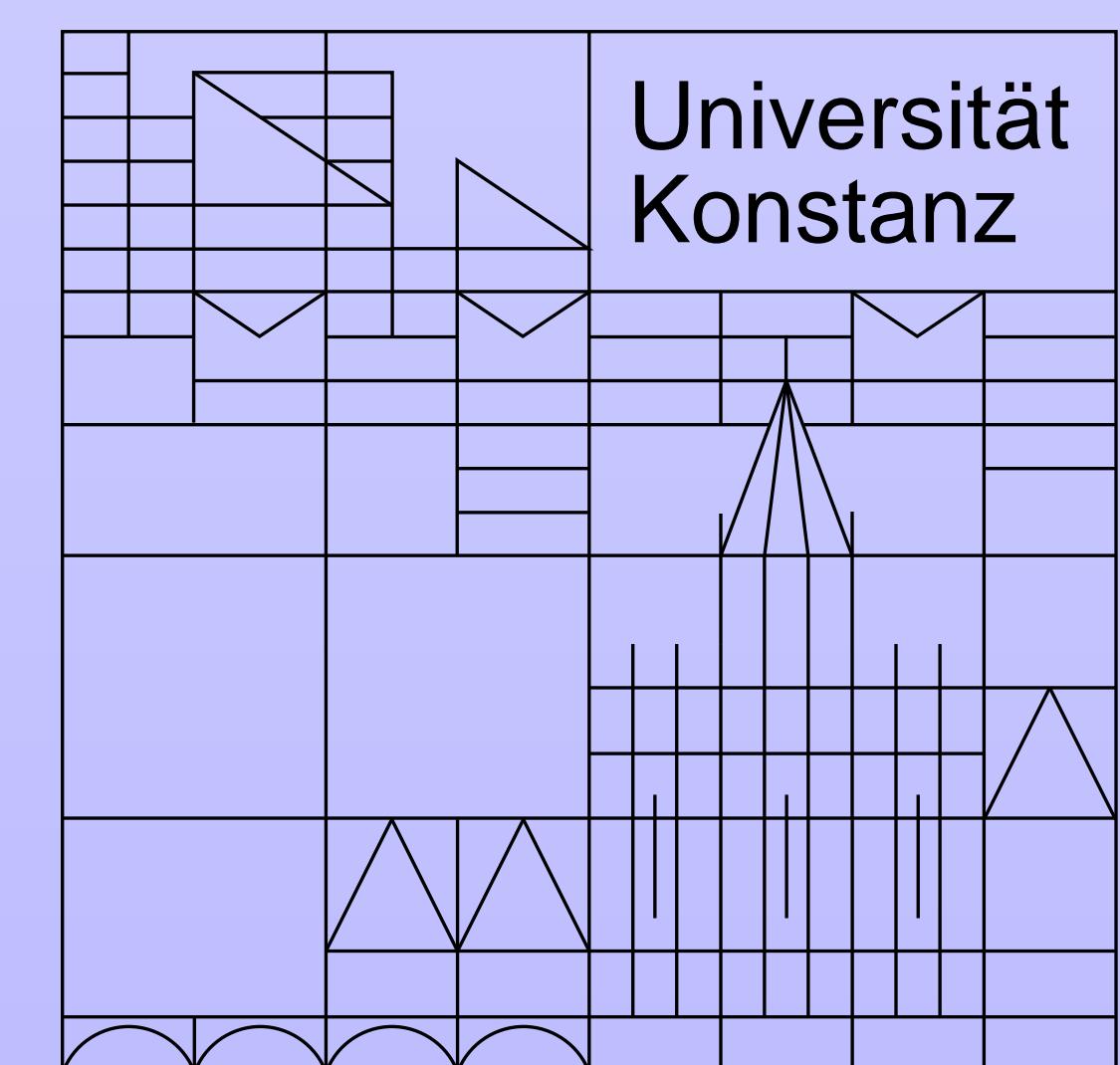


# Structural Model Complexes for 2-Oxoglutarate Dependent Iron Enzymes

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## 2-Oxoglutarate Dependent Iron Enzymes

In recent years protein structures of several 2-oxoglutarate dependent mononuclear non heme iron(II) oxygenases were resolved, such as deacetoxycephalosporin C synthase (DAOCS)<sup>[1]</sup> and taurine dioxygenase (TauD)<sup>[2]</sup>.

In the active site of these enzymes the iron(II) centre is coordinated by two histidines and one aspartate or glutamate, the so-called 2-His-1-carboxylate facial triad<sup>[3]</sup>. The 2-oxoglutarate co-substrate is bound via one carboxylate and the 2-oxo group to the iron center (Fig.1).

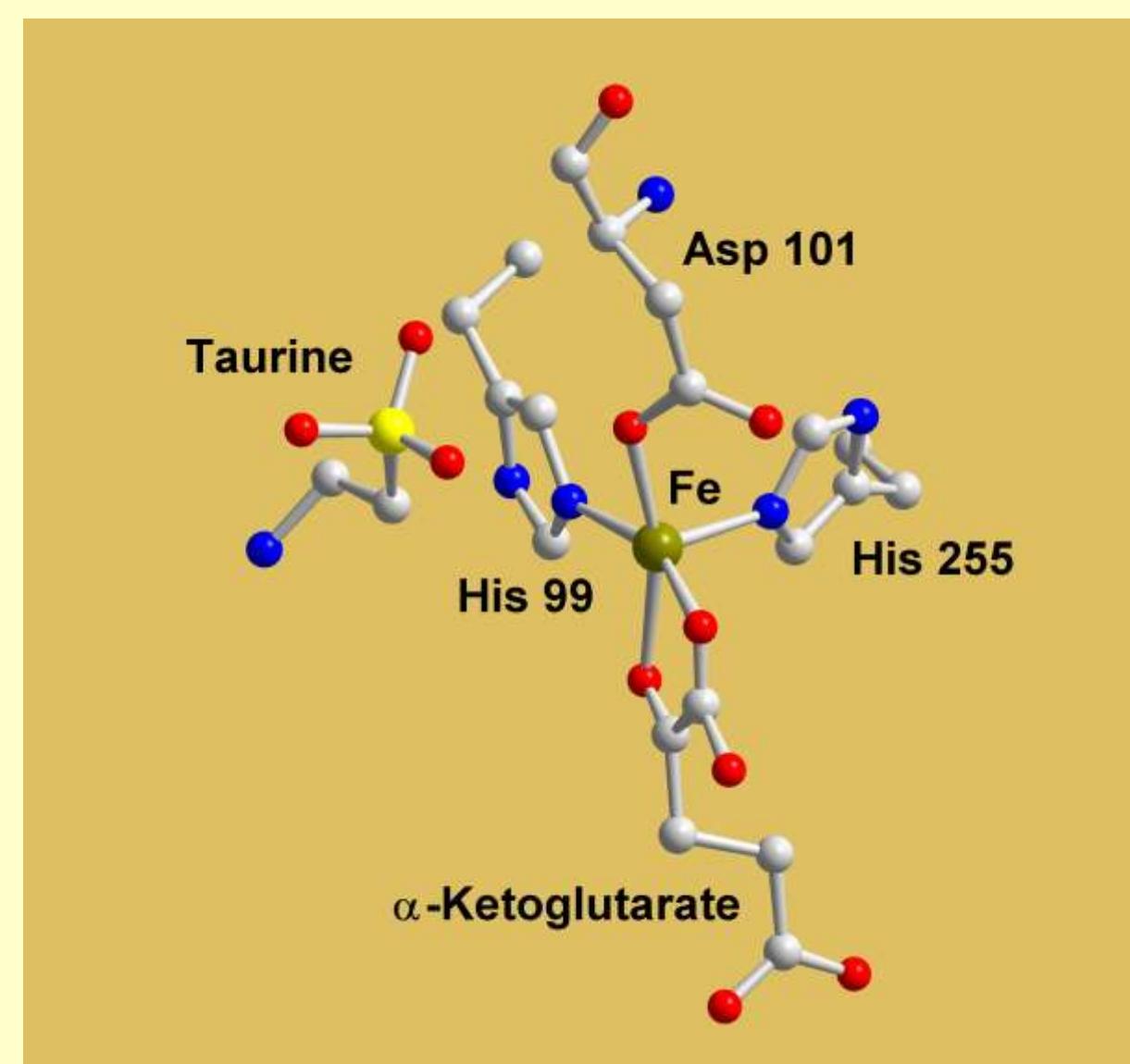
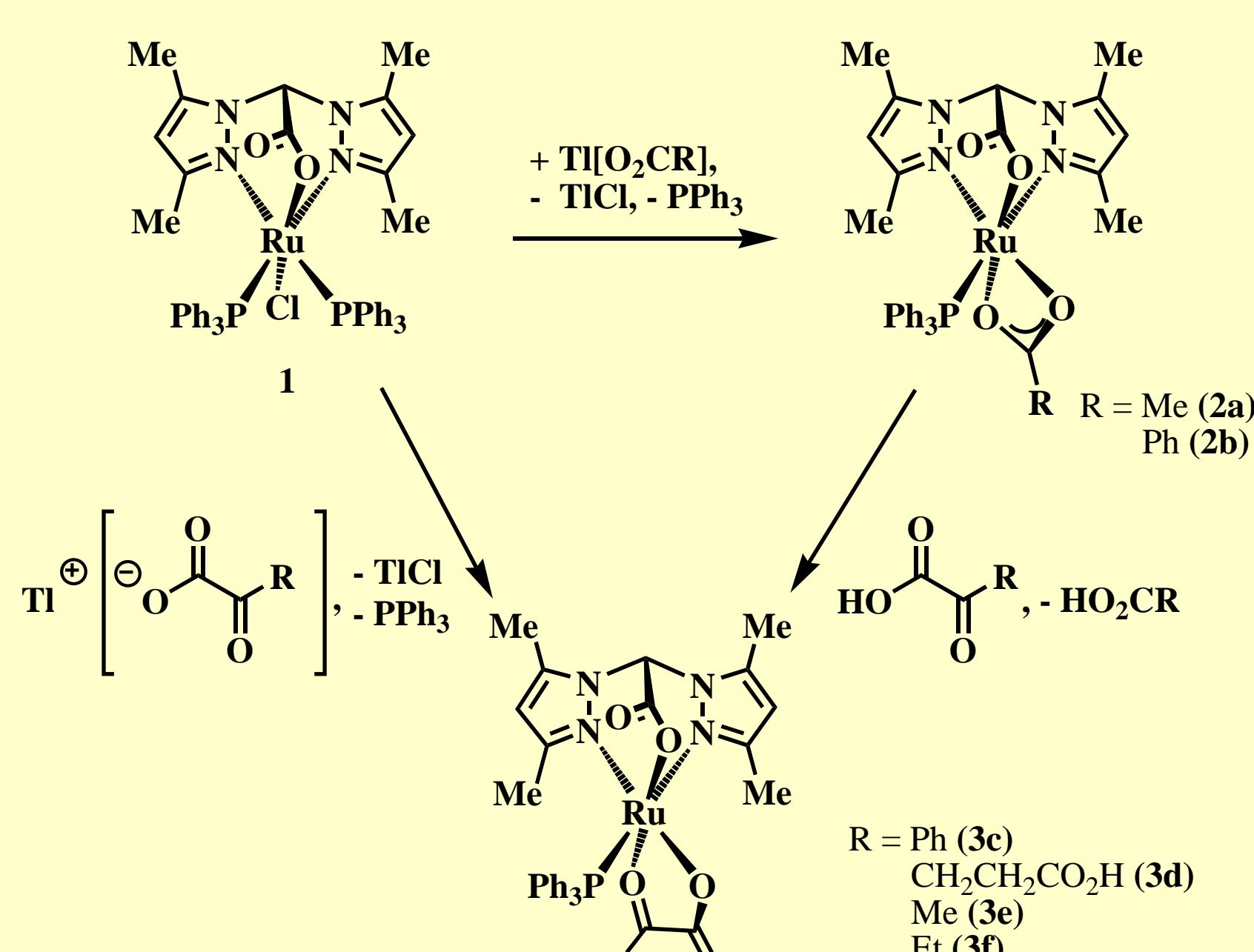


Fig.1: Active site of TauD with 2-oxoglutarate and taurine substrate (PDB-Code: 1GY9)<sup>[2]</sup>

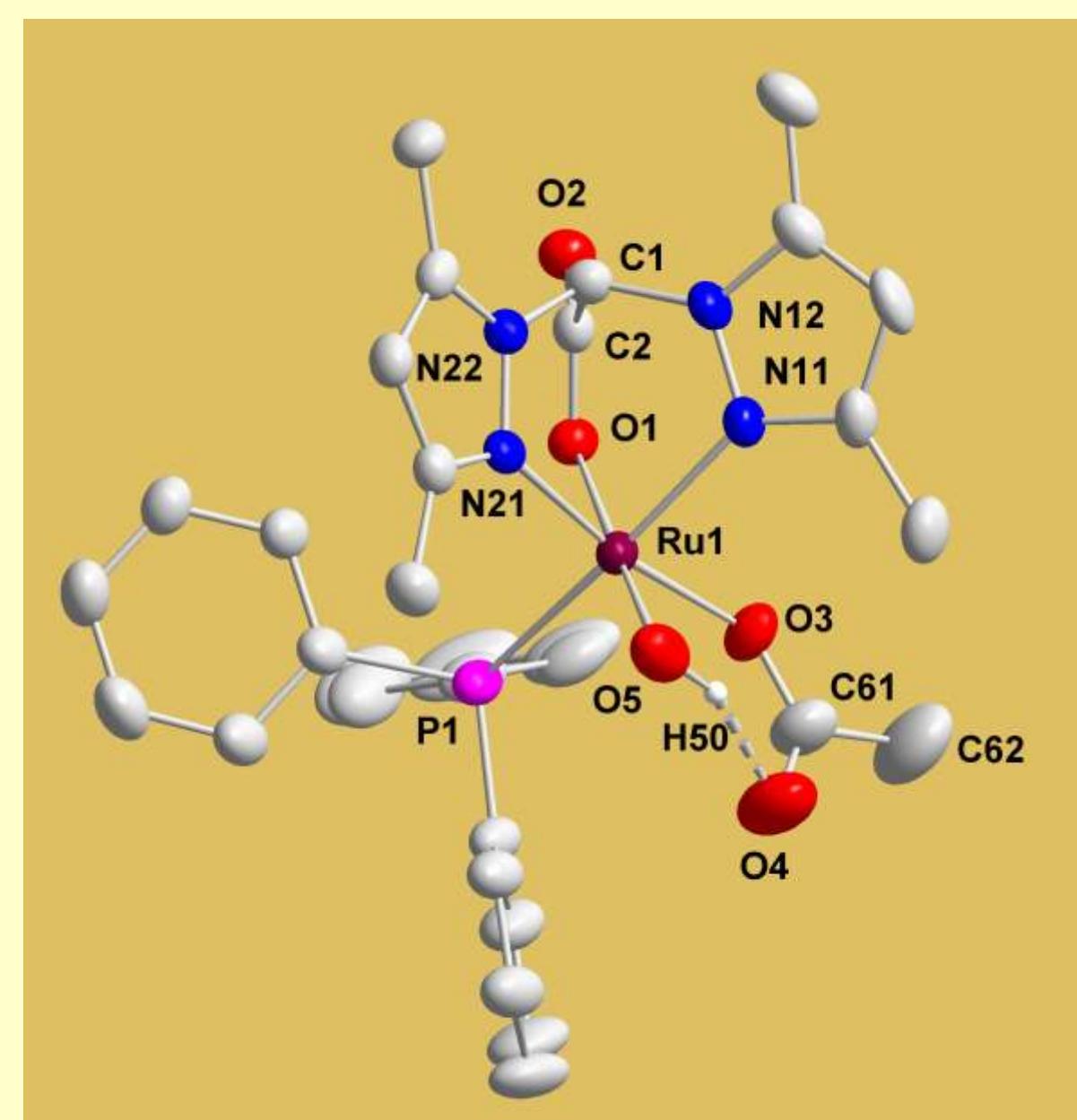


Fig.2: X-ray structure of [(bdmpza)Ru(H<sub>2</sub>O)(OAc)(PPh<sub>3</sub>)]

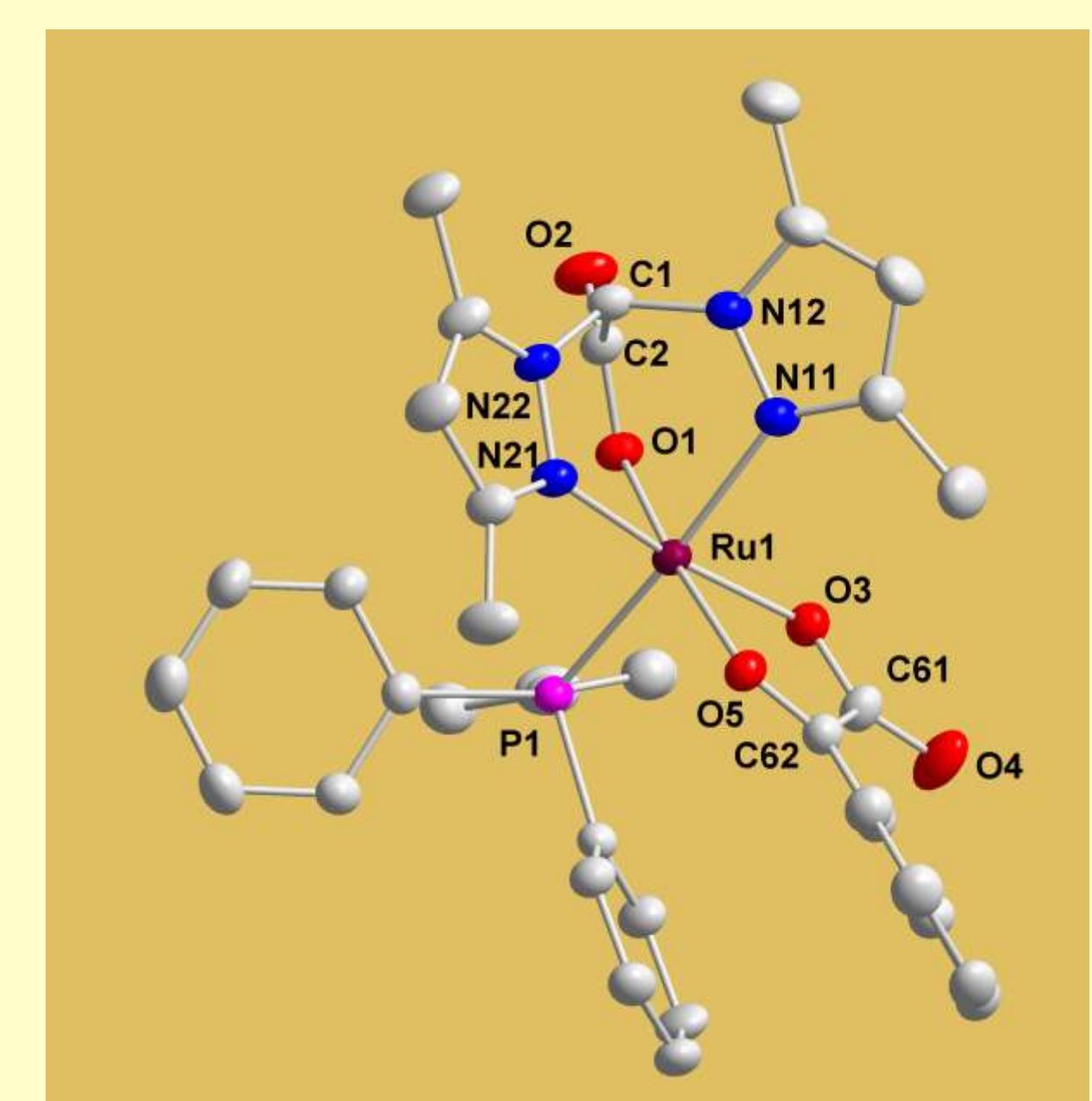
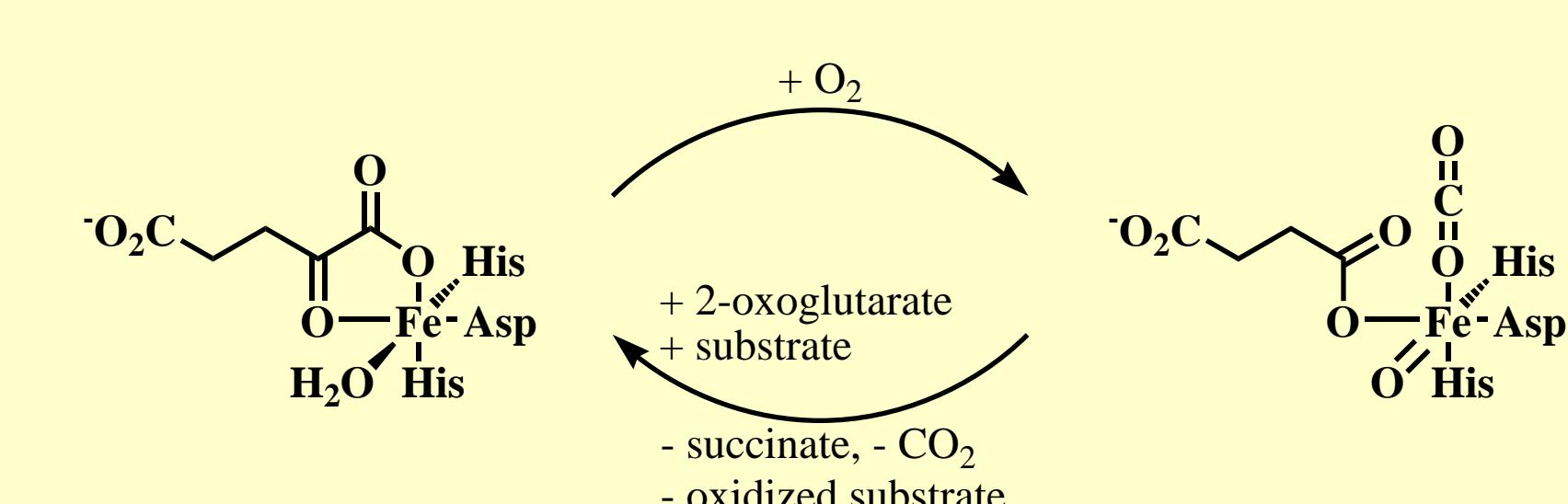


Fig.3: Molecular structure of [(bdmpza)Ru(O<sub>2</sub>CC(O)Ph)(PPh<sub>3</sub>)]

Carboxylato and 2-oxo carboxylato complexes are obtained by reaction of the ruthenium(II) complex [(bdmpza)RuCl(PPh<sub>3</sub>)<sub>2</sub>]<sup>[4]</sup> (bdmpza = bis(3,5-dimethylpyrazol-1-yl)acetate) with the corresponding thallium carboxylates (Scheme 1)<sup>[5]</sup>.

Reaction of the carboxylato complexes with 2-oxo acids also yields the 2-oxo carboxylato complexes. This reaction can be compared with the regeneration step of 2-oxoglutarate dependent enzymes (Scheme 2).

The coordination of the 2-oxo acid in the model complex is the same as in the enzyme (Fig. 1 & 3). The keto function binds *trans* to the carboxylate group of the aspartate or glutamate in the enzyme and *trans* to the carboxylate group of the bdmpza ligand in the model.



Scheme 2: Reaction cycle in 2-oxoglutarate dependent enzymes

## Iron Enzyme Inhibitors

The 2-oxoglutarate analogue *N*-oxalylglycine is a lead structure for inhibitors of enzymes such as prolyl 4-hydroxylase<sup>[6]</sup> and factor inhibiting HIF (FIH) (Fig.4)<sup>[7]</sup>. These inhibitors might be used in the therapy of rheumatoid arthritis and other fibrotic diseases<sup>[6]</sup>.

The 2-benzoyl-cyclohexane-1,3-diones type herbicides are potent inhibitors for the 4-hydroxyphenylpyruvate dioxygenase (HPPD). These triketones are also of pharmaceutical use for treatment of tyrosinaemia<sup>[8]</sup>.

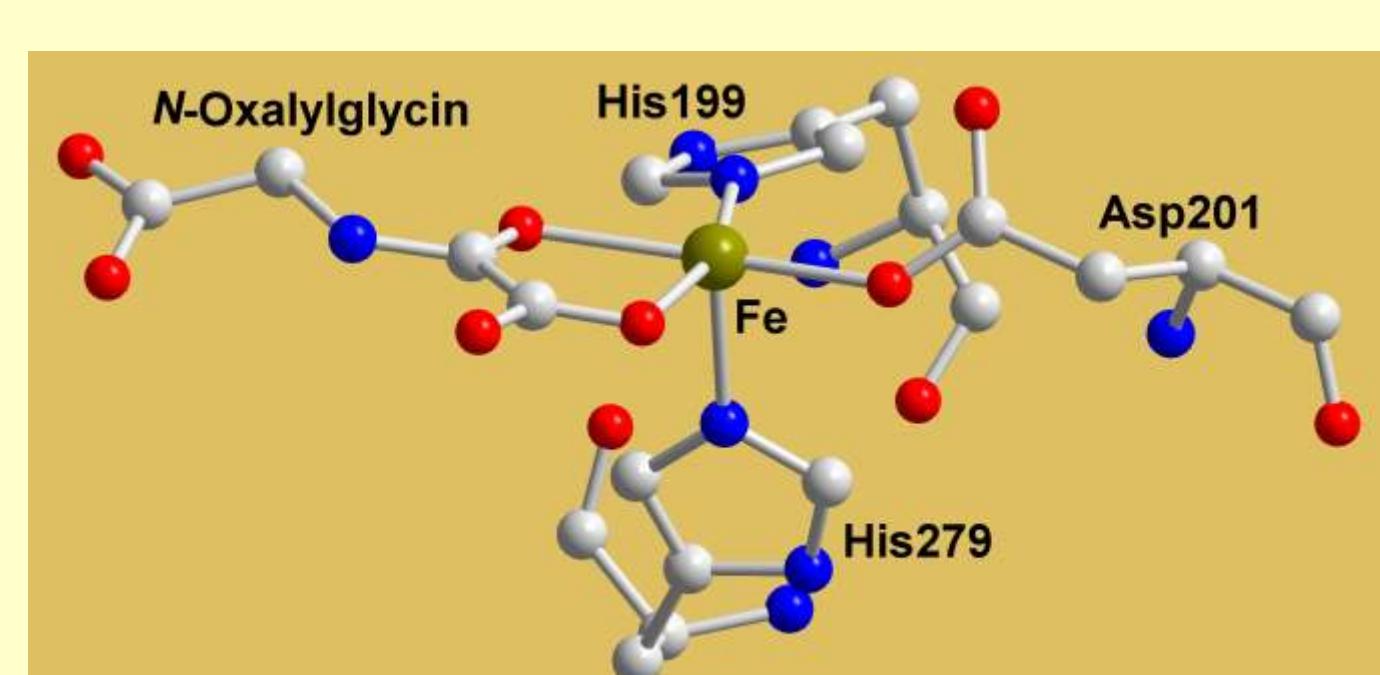
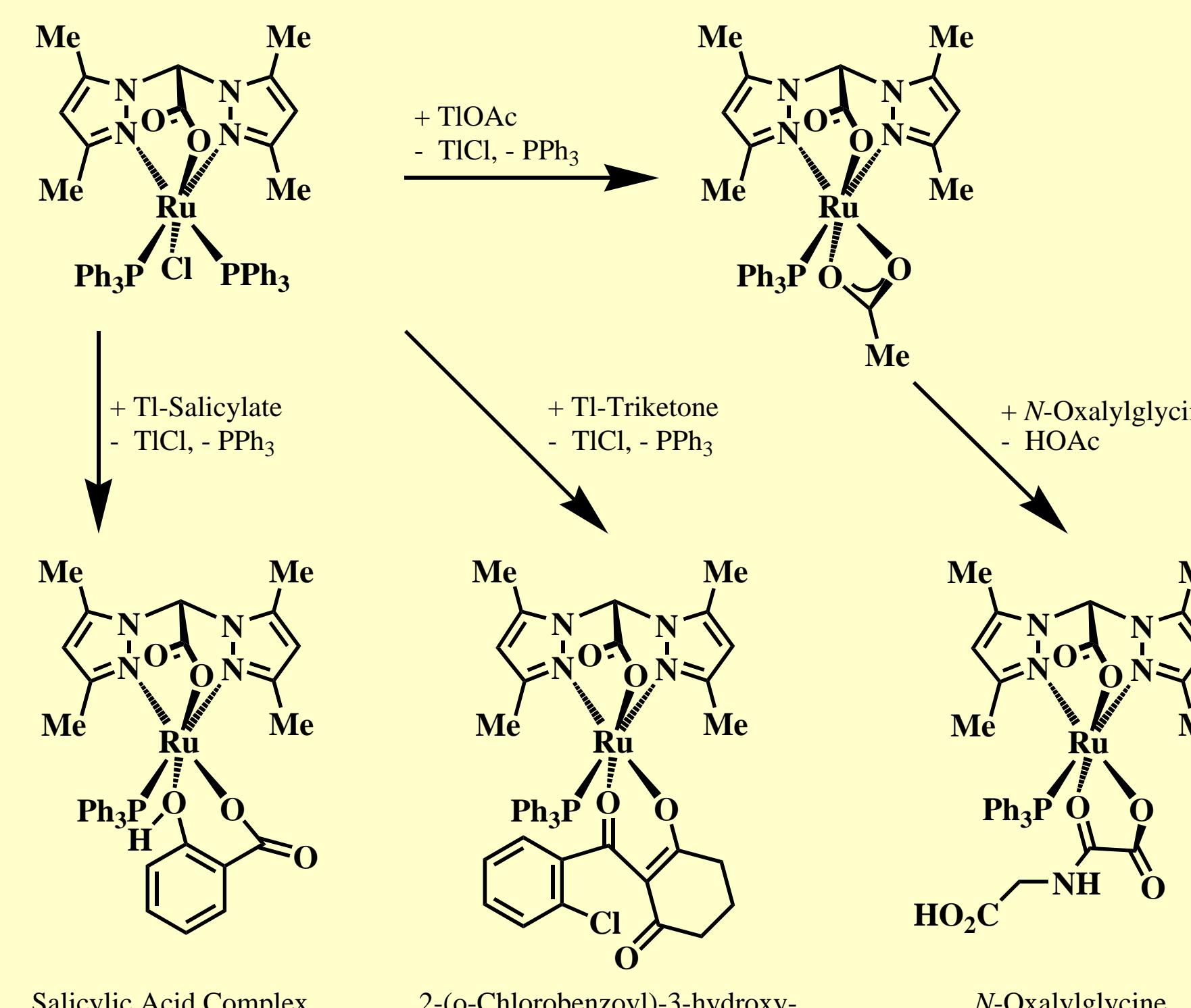


Fig.4: X-ray structure of the active site of FIH with *N*-oxalylglycine inhibitor (PDB-Code: 1H2K)<sup>[7]</sup>

We were able to obtain ruthenium model complexes with *N*-oxalylglycine, a triketone type inhibitor and salicylic acid (Scheme 3).



Scheme 3: Synthesis of inhibitor complexes

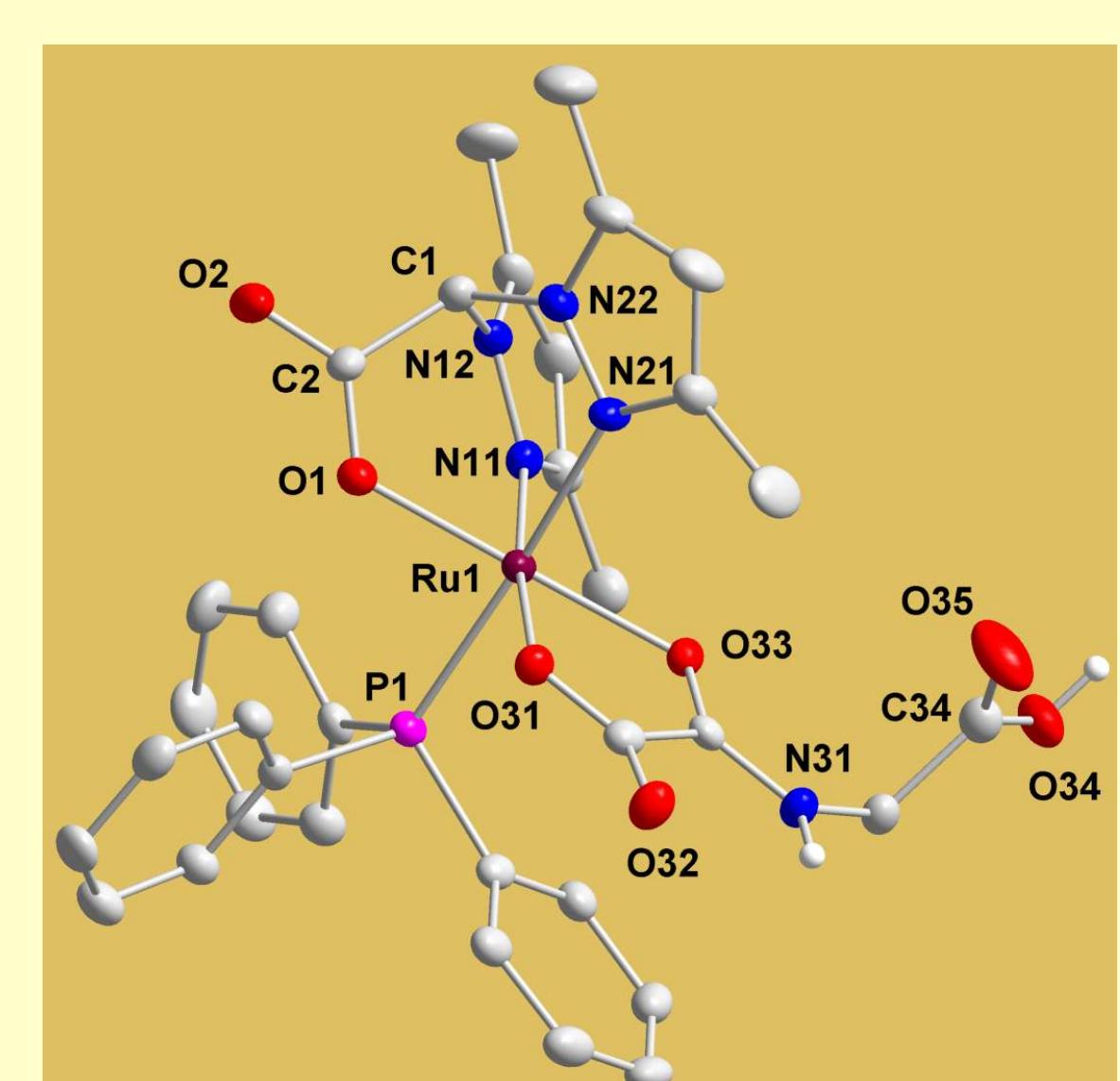
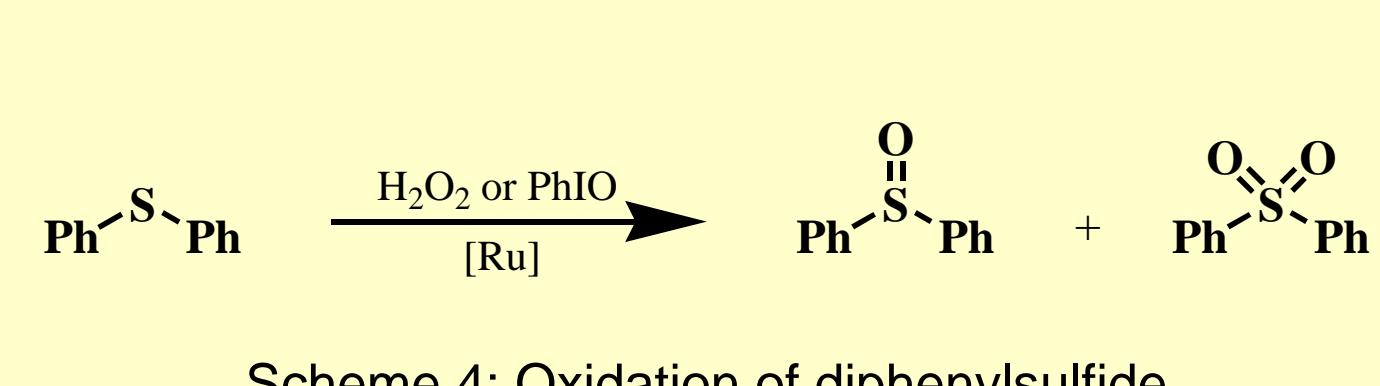


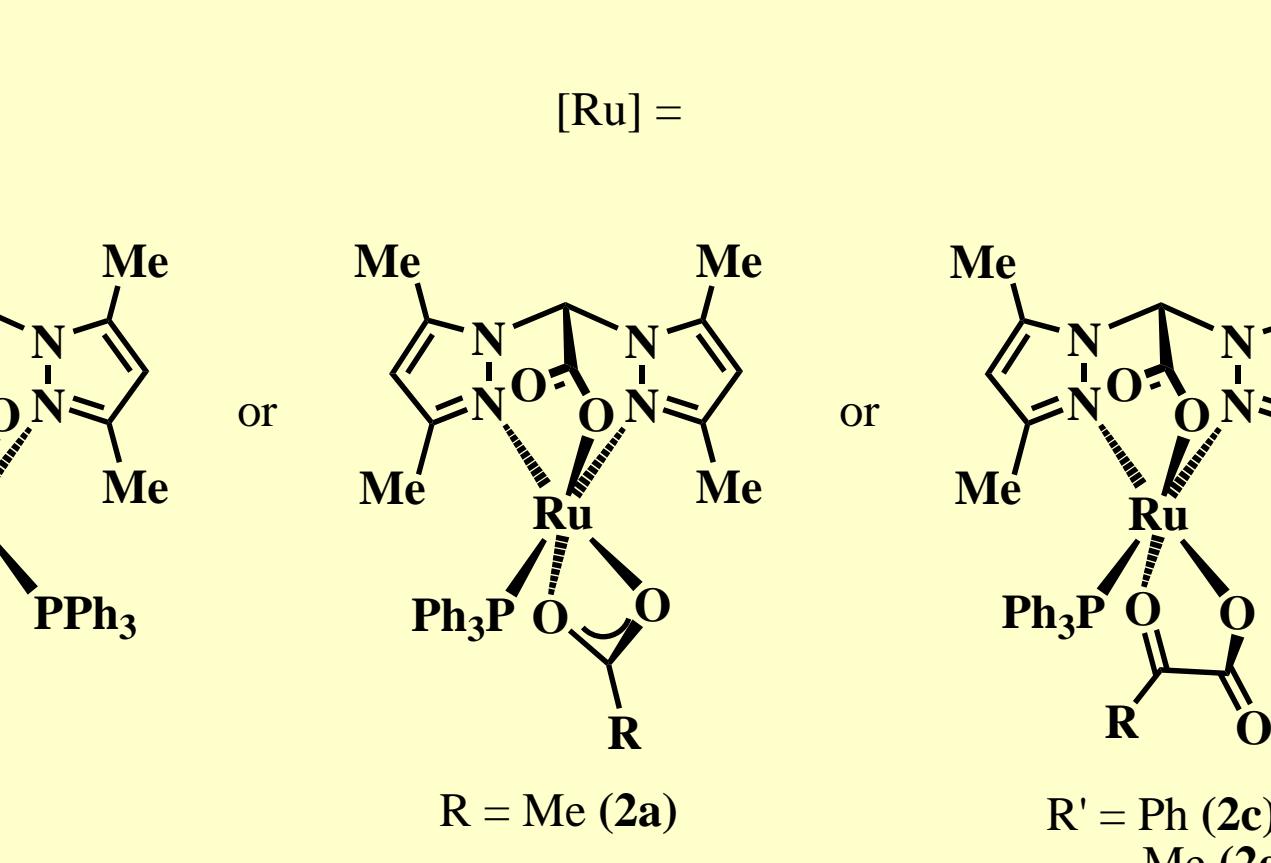
Fig.5: Molecular structure of a ruthenium model complex bearing *N*-oxalylglycine

## Oxidation Catalysis

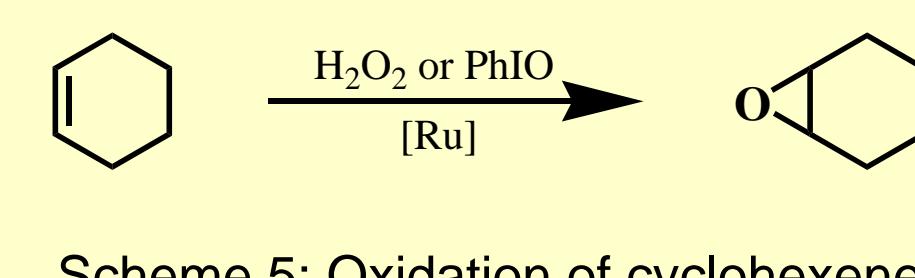
The ruthenium complexes 2a - 2f are good structural models for 2-oxoglutarate dependent enzymes. So far we observed no activation of molecular oxygen by these complexes (Scheme 6). Nevertheless, using oxidizing agents like H<sub>2</sub>O<sub>2</sub> or iodosobenzene they oxidize diphenylsulfide (Scheme 4) and cyclohexene (Scheme 5) in a bio-inspired “peroxide shunt” type reaction.



catalyst (10 mol%)	oxidizing agent	turnover	turnover number
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CCCH <sub>3</sub> ) 3a	H <sub>2</sub> O <sub>2</sub>	37.8	3.78
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CCCH <sub>3</sub> ) 3a	PhIO	75.3	7.53
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CC(O)Ph) 3c	H <sub>2</sub> O <sub>2</sub>	97.2	9.72
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CC(O)Ph) 3c	PhIO	69.8	6.98
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CC(O)Me) 3e	H <sub>2</sub> O <sub>2</sub>	21.7	2.17
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CC(O)Me) 3e	PhIO	51.7	5.17



Scheme 6: Catalytic active model complexes



catalyst (10 mol%)	oxidizing agent	turnover	turnover number
(bdmpza)Ru(PPh <sub>3</sub> ) <sub>2</sub> Cl 1	H <sub>2</sub> O <sub>2</sub>	-	-
(bdmpza)Ru(PPh <sub>3</sub> ) <sub>2</sub> Cl 1	PhIO	42.1	4.21
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CCCH <sub>3</sub> ) 3a	H <sub>2</sub> O <sub>2</sub>	37.8	3.78
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CCCH <sub>3</sub> ) 3a	PhIO	75.3	7.53
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CC(O)Ph) 3c	H <sub>2</sub> O <sub>2</sub>	97.2	9.72
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CC(O)Ph) 3c	PhIO	69.8	6.98
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CC(O)Me) 3e	H <sub>2</sub> O <sub>2</sub>	-	-
(bdmpza)Ru(PPh <sub>3</sub> )(O <sub>2</sub> CC(O)Me) 3e	PhIO	51.7	5.17

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